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HUABIO RESEARCH INC

Human/Monkey/Canine IL-1 beta ELISA Instructions

Cat:EH0006

Content

	CAT	Volume
❶ CP (Coated Plate)	EH0006CP	96 well
❷ S (Standard)	EH0006S	2 vial
❸ SD (Sample Diluent)	ESD01	15 ml/bottle
❹ DA (Detect Antibody)	EH0006DA	6 ml/bottle
❺ SH (Streptavidin-HRP)	ESH01	12 ml/bottle
❻ AB (Assay Buffer 1×)	EAB01	12 ml/bottle
❼ TS (TMB Substrate)	ETS01	12 ml/bottle
❽ SS (Stop Solution)	ESS01	12 ml/bottle
❾ WB (Wash Buffer 10×)	EWB01	50 ml/bottle
❿ SF (Sealer Film)	ESF01	6 pieces

NOTE: After the kit is opened, the stabilization period of each content is 30 days, so please use the kit within 30 days after opening.

REAGENT PREPARATION

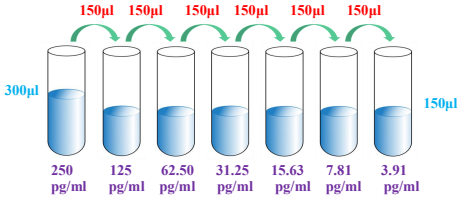
Standard Curve Preparation:

The **SD** (Sample Diluent) is used for serum and plasma. Other sample type, prepare the standard curve with whatever buffer (SPB, Sample Prepared Buffer) is used to prepare the sample, such as cell culture supernatant, tissue grinding liquid, cell lysate, etc. Urine sample use AB (Assay Buffer) prepare standard curve.

Reconstitute human IL-1 beta Standard by addition of distilled water as S. Reconstitution volume is stated on the label of the standard vial. Swirl or mix gently to insure complete and homogeneous solubilization (concentration of reconstituted standard = 2500 pg/ml).

Allow the standard to reconstitute for 10-30 minutes. Mix well prior to making dilutions.

The human IL-1 beta Standard EH0006S 2500 pg/ml 30 µl + 270 µl SPB serves as the high standard (250 pg/ml). Pipette 150 µl of SPB into each tube. Use the high standard to produce a 1:1 dilution series. Mix each tube thoroughly before the next transfer. SPB serves as the zero standard (0 pg/ml).



Washing Buffer (1×) Preparation

Pour entire contents (50 ml) of the **Washing Buffer Concentrate** (10×) into a clean 500 ml graduated cylinder. Bring to final volume of 500 ml with glass-distilled or deionized water. Transfer to a clean wash bottle and store at 2 to 25℃.

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use.

❶ Prepare all reagents and working standards as directed in the previous sections.

❷ Remove excess **CP** (Coated Plate) strips from the plate frame, return them to the foil pouch and reseal.

❸ Add 50 µl of **AB** (Assay Buffer) to each well.

❹ Add 50 µl or 10 µl of **Standard or sample** per well. Ensure reagent addition is uninterrupted and completed within 15 minutes.

❺ Add 50 µl of **DA** (Detect Antibody) to each well.

❻ Cover with an **SF** (Sealer Film). Incubate at room temperature (18 to 25℃) for 1 hour on a microplate **shaker** set at 500 rpm.

❼ Aspirate each well and **wash**, repeating the process four times. Wash by filling each well with WB (Washing Buffer 300 µl). Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining WB (Washing Buffer) by aspirating or decanting. Invert the plate and **blot** it against clean paper towels.

❽ Add 100 µl of **SH** (Streptavidin-HRP) to each well.

❾ Cover with a new **SF** (Sealer Film). Incubate at room temperature (18 to 25℃) for 30 min on a microplate **shaker** set at 500 rpm.

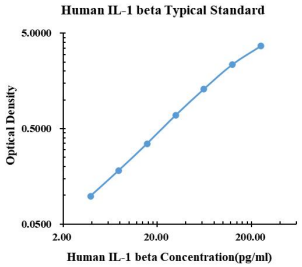
❿ Repeat aspiration/**wash** as in step 7.

⓫ Add 100 µl of **TS** (TMB Substrate) to each well. Incubate for 5-30 minutes at room temperature.

⓬ Add 100 µl of **SS** (Stop Solution) to each well.

⓭ Determine the optical density within 30 minutes, using microplate **reader** set to 450 nm corrected with 570 nm or 630 nm.

TYPICAL DATA



pg/ml	O.D.		Average	Corrected
0.00	0.0296	0.0307	0.0302	
3.91	0.1330	0.1232	0.1281	0.0980
7.81	0.2132	0.2122	0.2127	0.1826
15.63	0.3820	0.3760	0.3790	0.3489
31.25	0.7406	0.6980	0.7193	0.6892
62.50	1.3710	1.2880	1.3295	1.2994
125.00	2.4490	2.3060	2.3775	2.3474
250.00	3.7620	3.6230	3.6925	3.6624

SENSITIVITY

The minimum detectable dose (MDD) of human IL-1 beta is typically less than 0.09 pg/ml (50 µl of sample volume) or 0.18 pg/ml (10 µl of sample volume).

The MDD was determined by adding two standard deviations to the mean optical density value of ten zero standard replicates and calculating the corresponding concentration.

PRECISION

Intra-assay Precision (Precision within an assay) Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays)

	Intra-assay Precision			Inter-assay Precision		
Sample Number	S1	S2	S3	S1	S2	S3
	22	22	22	6	6	6
Average（pg/ml）	6.0	29.6	93.7	7.2	33.6	104.4
Standard Deviation	0.3	1.3	4.1	0.3	1.0	2.0
Coefficient of Variation（%）	4.8	4.5	4.4	4.1	2.9	1.9

RECOVERY

The spike recovery was evaluated by spiking 3 levels of human IL-1 beta into health human serum sample. The un-spiked serum was used as blank in this experiment.

The recovery ranged from 85% to 120% with an overall mean recovery of 95%.

LINEARITY

To assess the linearity of the assay, five samples were spiked with high concentration of IL-1 beta in human serum and diluted with Sample Diluent to produce samples with values within the dynamic range of the assay.

The linearity ranged from 89% to 118% with an overall mean recovery of 94%.

SAMPLE VALUES

Serum/Plasma – Thirty samples from apparently healthy volunteers, two samples from healthy monkeys and five samples from healthy canines were evaluated for the presence of human IL-1 beta in this assay. No medical histories were available for the donors.

Sample Matrix	Sample Evaluated	Range (pg/ml)	Detectable %	Mean of Detectable (pg/ml)
Human Serum	30	1.56-60.87	100	15.59
Monkey Serum	2	48.96-51.40	100	50.18
Canine Serum	5	12.00-16.03	100	14.42

n.d. = non-detectable. Samples measured below the sensitivity are considered to be non-detectable.