

HIV-1 p24 Instructions

Content

	CAT	Volume
 CP (Coated Plate) 	EH0078CP	96 well
8 (Standard)	EH0078S,S1~S7,S0	9 vial
SD (Sample Diluent)	ESD01	12 ml/bottle
4 AB (Assay Buffer 1×)	EAB01	6 ml/bottle
5 DA (Detect Antibody)	EH0078DA	6 ml/bottle
6 SH (Streptavidin-HRP)	ESH01	12 ml/bottle
TS (TMB Substrate)	ETS01	12 ml/bottle
8 SS (Stop Solution)	ESS01	12 ml/bottle
9 WB (Wash Buffer 10×)	EWB01	50 ml/bottle
Image: SF (Sealer Film)	ESF01	6 pieces

NOTE: After the kit is opened, the stabilization period of each content is 30 days, so please use the kit within 30 days after opening.

REAGENT PREPARATION

Washing Buffer (1×) Preparation

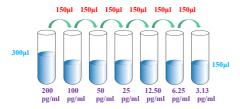
Pour entire contents (50 ml) of the Washing Buffer Concentrate (10×) into a clean 500 ml graduated cylinder. Bring to final volume of 500 ml with glass-distilled or deionized water. Transfer to a clean wash bottle and store at 2 to 25°C.

Standard Curve Preparation:

S1 to S7 and S0 is ready to use for serum, plasma and cell culture supernatant.

Other sample type, prepare the standard curve with whatever buffer (SPB, Sample Prepared Buffer) is used to prepare the sample, such as tissue grinding liquid, cell lysate, etc. Urine sample use AB (Assay Buffer) prepare standard curve.

The HIV-1 p24 Standard EH0078S 2000 pg/ml $30 \ \mu$ l + 270 μ l SPB serves as the high standard (5000 pg/ml). Pipette 150 μ l of SPB into each tube. Use the high standard to produce a 1:1 dilution series. Mix each tube thoroughly before the next transfer. SPB serves as the zero standard (0 pg/ml).



Cat: EH0078

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use.

 Prepare all reagents and working standards as directed in the previous sections.

Remove excess CP (Coated Plate) strips from the plate frame, return them to the foil pouch and reseal.

3 Add 50 μl of AB (Assay Buffer) to each well.

4 Add 50 μl or 10 μl of Standard or sample per well. Ensure reagent addition is uninterrupted and completed within 15 minutes.

(5) Add 50 μ l of DA (Detect Antibody) to each well.

6 Cover with an SF (Sealer Film). Incubate at room temperature (18 to 25°C) for 1 hours on a microplate shaker set at 500 rpm.

Aspirate each well and wash, repeating the process four times. Wash by filling each well with WB (Washing Buffer 300 μl). Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining WB (Washing Buffer) by aspirating or decanting. Invert the plate and blot it against clean paper towels.

8 Add 100 μ l of SH (Streptavidin-HRP) to each well.

Over with a new SF (Sealer Film). Incubate at room temperature (18 to 25°C) for 30 min on a microplate shaker set at 500 rpm.

10 Repeat aspiration/wash as in step 7.

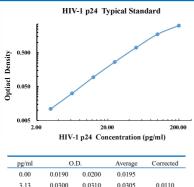
Add 100 μl of TS (TMB Substrate) to each well. Incubate for 5-30 minutes at room temperature.

12 Add 100 μl of SS (Stop Solution) to each well.

B Determine the optical density within 30 minutes, using microplate reader set to 450 nm corrected with 570 nm or 630 nm.



TYPICAL DATA



0.00	0.0190	0.0200	0.0195		
3.13	0.0300	0.0310	0.0305	0.0110	
6.25	0.0520	0.0500	0.0510	0.0315	
12.50	0.1150	0.1140	0.1145	0.0950	
25.00	0.2920	0.2800	0.2860	0.2665	
50.00	0.7590	0.7000	0.7295	0.7100	
100.00	1.8220	1.7240	1.7730	1.7535	
200.00	3.2790	3.0910	3.1850	3.1655	

SENSITIVITY

The minimum detectable dose (MDD) of HIV-1 p24 is typically less than 0.64 pg/ml (50 μ l of sample volume) or 2.62 pg/ml (10 μ l of sample volume).

The MDD was determined by adding two standard deviations to the mean optical density value of ten zero standard replicates and calculating the corresponding concentration.

PRECISION

■ Intra-assay Precision (Precision within an assay) Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays)

	Intra-assay Precision			Inter-assay Precision			
Sample Number	S1	S2	83		S1	S2	\$3
	22	22	22		6	6	6
Average (pg/ml)	4.6	18.4	58.9		4.2	19.5	59.7
Standard deviation	0.12	0.59	4.1		0.21	0.72	3.8
Coefficient of variation (%)	2.6	3.2	6.9		5.0	3.7	6.4

RECOVERY

The spike recovery was evaluated by spiking 3 levels of HIV-1 p24 into serum sample. The un-spiked serum was used as blank in this experiment.

The recovery ranged from 94% to 105% with an overall mean recovery of 98%.

LINEARITY

To assess the linearity of the assay, five samples were spiked with high concentration of serum sample and diluted with Sample Diluent to produce samples with values within the dynamic range of the assay.

The linearity ranged from 95% to 104% with an overall mean recovery of 99%.

Sample Diluent

If the concentration of the sample is too high, the sample can be diluted with Sample Diluent.