

# Human CCL16/LEC ELISA Instructions

**Cat:EHY0294**

## Content

|                              | CAT             | Volume       |
|------------------------------|-----------------|--------------|
| ① CP (Coated Plate)          | EHY0294CP       | 96 well      |
| ② S (Standard)               | EHY0294S1-S7,S0 | 8 vial       |
| ③ DA-H (Detect Antibody-HRP) | EHY0294DA-H     | 6 ml/bottle  |
| ④ AB (Assay Buffer 1×)       | EAB01           | 12 ml/bottle |
| ⑤ TS (TMB Substrate)         | ETS01           | 12 ml/bottle |
| ⑥ SS (Stop Solution)         | ESS01           | 12 ml/bottle |
| ⑦ WB (Wash Buffer 10×)       | EWB01           | 50 ml/bottle |
| ⑧ SF (Sealer Film)           | ESF01           | 6 pieces     |

**NOTE:** After the kit is opened, the stabilization period of each content is 30 days, so please use the kit within 30 days after opening.

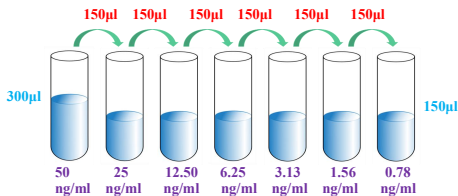
## REAGENT PREPARATION

### Washing Buffer (1×) Preparation

Pour entire contents (50 ml) of the **Washing Buffer Concentrate** (10×) into a clean 500 ml graduated cylinder. Bring to final volume of 500 ml with glass-distilled or deionized water. Transfer to a clean wash bottle and store at 2 to 25°C.

### Standard Curve Preparation:

S1 to S7 and S0 is ready to use for serum and plasma.



## ASSAY PROCEDURE

**Bring all reagents and samples to room temperature before use.**

① Prepare all reagents and working standards as directed in the previous sections.

② Remove excess **CP** (Coated Plate) strips from the plate frame, return them to the foil pouch and reseal.

③ Add 50 µl of **AB** (Assay Buffer) to each well.

④ Add 10 µl of **Standard or sample** per well.

Ensure reagent addition is uninterrupted and completed within 15 minutes.

⑤ Add 50 µl of **DA-H** (Detect Antibody-HRP) to each well.

⑥ Cover with an **SF** (Sealer Film). Incubate at room temperature (18 to 25°C) for 30 min on a microplate **shaker** set at 500 rpm.

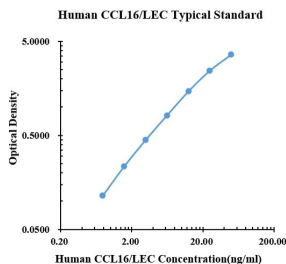
⑦ Aspirate each well and **wash**, repeating the process four times. Wash by filling each well with **WB** (Washing Buffer 300 µl). Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining **WB** (Washing Buffer) by aspirating or decanting. Invert the plate and **blot** it against clean paper towels.

⑧ Add 100 µl of **TS** (TMB Substrate) to each well. Incubate for 5-30 minutes at room temperature.

⑨ Add 100 µl of **SS** (Stop Solution) to each well.

⑩ Determine the optical density within 30 minutes, using microplate **reader** set to 450 nm corrected with 570 nm or 630 nm.

## TYPICAL DATA



| ng/ml | O.D.   | Average | Corrected |
|-------|--------|---------|-----------|
| 0.00  | 0.0054 | 0.0054  |           |
| 0.78  | 0.1235 | 0.1153  | 0.1194    |
| 1.56  | 0.2368 | 0.2407  | 0.2334    |
| 3.13  | 0.4504 | 0.4531  | 0.4464    |
| 6.25  | 0.8144 | 0.8275  | 0.8210    |
| 12.50 | 1.4330 | 1.5100  | 1.4715    |
| 25.00 | 2.4360 | 2.4490  | 2.4371    |
| 50.00 | 3.5390 | 3.6880  | 3.6135    |

## SENSITIVITY

The minimum detectable dose (MDD) of human CCL16/LEC is typically less than 0.01 ng/ml (10  $\mu$ l of sample volume).

The MDD was determined by adding two standard deviations to the mean optical density value of ten zero standard replicates and calculating the corresponding concentration.

## PRECISION

**Intra-assay Precision (Precision within an assay)** Three samples of known concentration were tested twenty-two times on one plate to assess intra-assay precision.

**Inter-assay Precision (Precision between assays)**

|                              | Intra-assay Precision |     |      | Inter-assay Precision |     |      |
|------------------------------|-----------------------|-----|------|-----------------------|-----|------|
| Sample Number                | S1                    | S2  | S3   | S1                    | S2  | S3   |
|                              | 22                    | 22  | 22   | 6                     | 6   | 6    |
| Average (ng/ml)              | 1.3                   | 6.9 | 17.2 | 1.5                   | 6.4 | 16.2 |
| Standard Deviation           | 0.1                   | 0.3 | 0.8  | 0.1                   | 0.3 | 0.6  |
| Coefficient of Variation (%) | 4.3                   | 4.8 | 4.5  | 4.1                   | 4.7 | 3.4  |

## RECOVERY

The spike recovery was evaluated by spiking 3 levels of human CCL16/LEC into health human serum sample. The un-spiked serum was used as blank in this experiment.

The recovery ranged from 80% to 109% with an overall mean recovery of 93%.

## LINEARITY

To assess the linearity of the assay, five samples were spiked with high concentration of CCL16/LEC in human serum and diluted with Sample Diluent to produce samples with values within the dynamic range of the assay.

The linearity ranged from 80% to 103% with an overall mean recovery of 92%.

## SAMPLE VALUES

Serum/Plasma – Thirty samples from apparently healthy volunteers were evaluated for the presence of human CCL16/LEC in this assay. No medical histories were available for the donors.

| Sample Matrix | Sample Evaluated | Range (ng/ml) | Detectable % | Mean of Detectable (ng/ml) |
|---------------|------------------|---------------|--------------|----------------------------|
| Serum         | 30               | 4.33-13.11    | 100          | 8.66                       |

n.d. = non-detectable. Samples measured below the sensitivity are considered to be non-detectable.