

# **Mouse CRP/C-Reactive Protein ELISA Instructions**

# CONTENT

	CAT	Volume
<ol> <li>CP (Coated Plate)</li> </ol>	EM0035CP	96 well
8 (Standard)	EM0035S1	2 vial
3 DA-H (Detect Antibody-HRP)	EM0035DA-H	6 ml/bottle
4 SD (Sample Diluent)	ESD01	15 ml/bottle
6 AB (Assay Buffer 1×)	EAB01	12 ml/bottle
6 TS (TMB Substrate)	ETS01	12 ml/bottle
SS (Stop Solution)	ESS01	12 ml/bottle
8 WB (Wash Buffer 10×)	EWB01	50 ml/bottle
9 SF (Sealer Film)	ESF01	6 pieces

**NOTE:** After the kit is opened, the stabilization period of each content is 30 days.

### SAMPLE DILUTION

Samples such as serum  $\smallsetminus$  plasma require at least a 200-fold dilution into Sample Diluent. A suggested 200-fold dilution is 5  $\mu l$  of sample + 995  $\mu l$  of Sample Diluent.

# **REAGENT PREPARATION**

#### Washing Buffer (1×) Preparation

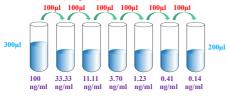
Pour entire contents (50 ml) of the Washing Buffer Concentrate (10x) into a clean 500 ml graduated cylinder. Bring to final volume of 500 ml with glass-distilled or deionized water. Transfer to a clean wash bottle and store at 2 to 25°C.

#### Standard Curve Preparation:

Reconstitute Mouse CRP Standard by addition of distilled water as S1. Reconstitution volume is stated on the label of the standard vial. Swirl or mix gently to insure complete and homogeneous solubilization (concentration of reconstituted standard = 100 ng/ml).

Allow the standard to reconstitute for 10-30 minutes. Mix well prior to making dilutions.

Pipette 200  $\mu$ l of Sample Diluent into each tube. Use the high standard to produce a 1:2 dilution series. Mix each tube thoroughly before the next transfer. Sample Diluent serves as the zero standard (0 ng/ml).



# Cat: EM0035

# ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use.

 Prepare all reagents and working standards as directed in the previous sections.

2 Remove excess CP (Coated Plate) strips from the plate frame, return them to the foil pouch and reseal.

3 Add 50 μl of AB (Assay Buffer) to each well.

4 Add 50 μl of Standard or sample per well. Ensure reagent addition is uninterrupted and completed within 15 minutes.

**5** Cover with an SF (Sealer Film). Incubate at room temperature (18 to 25°C) for 30 minutes on a microplate shaker set at 500 rpm.

6 Aspirate each well and wash, repeating the process four times. Wash by filling each well with WB (Washing Buffer 300 μl). Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining WB (Washing Buffer) by aspirating or decanting. Invert the plate and blot it against clean paper towels.

Add 50 μl of DA-H (Detect Antibody-HRP) to each well.

Over with a new SF (Sealer Film). Incubate at room temperature (18 to 25°C) for 10 minutes on a microplate shaker set at 500 rpm.

9 Repeat aspiration/wash as in step 6.

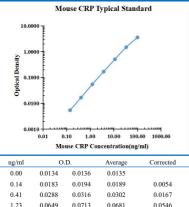
Model 100 μl of TS (TMB Substrate) to each well. Incubate for 5-30 minutes at room temperature.

1 Add 100 μl of SS (Stop Solution) to each well.

Determine the optical density within 30 minutes, using microplate reader set to 450 nm corrected with 570 nm or 630 nm.



# **TYPICAL DATA**



0.11	0.0200	0.0510	0.0502	0.0107
1.23	0.0649	0.0713	0.0681	0.0546
3.70	0.1768	0.1872	0.1820	0.1685
11.11	0.5001	0.5427	0.5214	0.5079
33.33	1.4370	1.5650	1.5010	1.4875
100.00	3.5510	3.6270	3.5890	3.5755

# **SENSITIVITY**

The minimum detectable dose (MDD) of mouse CRP is typically less than 0.01 ng/ml.

The MDD was determined by adding two standard deviations to the mean optical density value of ten zero standard replicates and calculating the corresponding concentration.

# PRECISION

Intra-assay Precision (Precision within an assay) Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays)

	Intra-assay Precision			Inter-assay Precision		
Sample	<b>S</b> 1	S2	S3	<b>S</b> 1	S2	S3
Number	22	22	22	6	6	6
Average (ng/ml)	1.6	8.4	30.4	2.0	10.0	32.0
Standard Deviation	0.1	0.3	1.1	0.0	0.2	0.6
Coefficient of Variation (%)	3.6	3.0	3.7	1.4	1.6	1.8

# RECOVERY

The spike recovery was evaluated by spiking 3 levels of mouse CRP into health mouse serum sample. The un-spiked serum was used as blank in this experiment.

The recovery ranged from 103% to 136% with an overall mean recovery of 125%.

# LINEARITY

To assess the linearity of the assay, five samples were spiked with high concentration of CRP in mouse serum and diluted with Sample Diluent to produce samples with values within the dynamic range of the assay.

The linearity ranged from 90% to 125% with an overall mean recovery of 98%.

# SAMPLE VALUES

Serum/Plasma – Thirty samples from apparently healthy mice were evaluated for the presence of mouse CRP in this assay. No medical histories were available for the donors.

Sample Matrix	Sample Evaluated	Range (ng/ml)	Detectable %	Mean of Detectable (ng/ml)
Serum	30	25.47-93.93	100	64.56

n.d. = non-detectable. Samples measured below the sensitivity are considered to be non-detectable.